

SuperScript[™] II Reverse Transcriptase

Cat. No. 18064-022 Cat. No. 18064-014 Cat. No. 18064-071 Size: 2,000 units Size: 10,000 units Size: 4 × 10,000 units Store at -20°C (non-frost-free)

Conc. 200 U/µL

Description

SuperScript[™] II Reverse Transcriptase (RT) is an engineered version of M-MLV RT with reduced RNase H activity and increased thermal stability. The enzyme is purified to near homogeneity from *E. coli* containing the modified *pol* gene of Moloney Murine Leukemia Virus (1,2). The enzyme can be used to synthesize first-strand cDNA at higher temperatures than conventional M-MLV RT, providing increased specificity, higher yields of cDNA, and more full-length product. It can generate cDNA up to 12.3 kb.

Components

SuperScript[™] II RT, 5X First-Strand Buffer (250 mM Tris-HCl, pH 8.3 at room temperature; 375 mM KCl; 15 mM MgCl₂), 0.1 M DTT

Storage Buffer

20 mM Tris-HCl (pH 7.5), 100 mM NaCl, 0.1 mM EDTA, 1 mM DTT, 0.01% (v/v) NP-40, 50% (v/v) glycerol

Storage Conditions

Store all components at -20° C in a non-frost-free freezer. Thaw 5X First-Strand Buffer and 0.1 M DTT at room temperature just prior to use and refreeze immediately.

Unit Definition

One unit incorporates 1 nmole of dTTP into acid-precipitable material in 10 min. at 37° C using poly(A)•oligo(dT)₂₅ as template-primer (3).

Intended Use

For research use only. Not intended for any animal or human therapeutic or diagnostic use.

Part no. 18064.pps

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First-Strand cDNA Synthesis Using SuperScript[™] II RT

A 20- μL reaction volume can be used for 1 ng–5 μg of total RNA or 1–500 ng of mRNA.

1.	1. Add the following components to a nuclease-free microcentrifuge tube:		
	Oligo(dT) ₁₂₋₁₈ (500 µg/mL) or	1 μL	
	50–250 ng random primers or		
	2 pmole gene-specific primer (GSP)		
	1 ng to 5 μg total RNA <i>or</i>	x μL	
	1–500 ng of mRNA		
	1 μL dNTP Mix (10 mM each)	1 μL	
	Sterile, distilled water	to 12 μL	
2.	Heat mixture to 65°C for 5 min and quick chill on ice. Collect the contents of the tube by brief centrifugation and add:		
	5X First-Strand Buffer	4 µL	
	0.1 M DTT	2 μL	
	RNaseOUT TM (40 units/ μ L) (optional)*	1 μL	
	*RNaseOUT [™] (Cat. No. 10777-019) is required if using <50 ng starting RNA		
3.	Mix contents of the tube gently. If you are using oligo(dT) ₁₂₋₁₈ or GSP.		

- Mix contents of the tube gently. If you are using oligo(dT)₁₂₋₁₈ or GSP, incubate at 42°C for 2 min. If you are using random primers, incubate at 25°C for 2 min.
- Add 1 µL (200 units) of SuperScript[™] II RT and mix by pipetting gently up and down. If you are using less than 1 ng of RNA, reduce the amount of

SuperScript^{\mathbb{M}} II RT to 0.25 μ L (50 units) and add sterile, distilled water to a 20 μ L final volume.

If you are using random primers, incubate tube at 25°C for 10 min.

- 5. Incubate at 42°C for 50 min.
- 6. Inactivate the reaction by heating at 70°C for 15 min.

First-Strand cDNA Synthesis Using SuperScript[™] II RT, Continued The cDNA can now be used as a template for amplification in PCR. However, amplification of some PCR targets (>1 kb) may require the removal of RNA complementary to the cDNA. To remove RNA complementary to the cDNA, add 1 µL (2 units) of *E. coli* RNase H and incubate at 37°C for 20 min.

PCR

The following is intended as a guideline and starting point when using firststrand cDNA in PCR with *Taq* DNA polymerase. The optimal concentration of Mg⁺⁺ will vary depending on the template and primer pair.

Use only 10% of the first-strand reaction for PCR. Higher volumes may not increase amplification and may result in decreased amounts of PCR product.

1. Add the following to a PCR tube:

10X PCR Buffer [200 mM Tris-HCl (pH 8.4), 500 mM KCl]	5 µL
50 mM MgCl ₂	1.5 μL
10 mM dNTP Mix	1 μL
Forward primer (10 μM)	1 μL
Reverse primer (10 µM)	1 μL
Taq DNA polymerase (5 U/ μ L)	0.4 μL
cDNA from first-strand reaction	2 μL
autoclaved, distilled water	to 50 µL

- Mix gently and layer with 1–2 drops (~50 μL) of silicone oil. (Note: silicone oil is unnecessary in thermal cyclers equipped with a heated lid.)
- 3. Heat reaction to 94°C for 2 min to denature.
- 4. Perform 15 to 40 cycles of PCR. Use the recommended annealing and extension conditions for your *Taq* DNA polymerase.

Product Qualification

The Certificate of Analysis provides detailed quality control information for each product. Certificates of Analysis are available at www.invitrogen.com/support.

Additional Products

RNaseOUT^m Recombinant Ribonuclease Inhibitor (40 units/ μ L) is available separately from Invitrogen (Cat. no. 10777-019).

References

- Kotewicz, M.L., D'Alessio, J.M., Driftmier, K.M., Blodgett, K.P., and Gerard, G.F. (1985) Gene 35, 249.
- Gerard, G.F., D'Alessio, J.M., Kotewicz, M.L., and Noon, M.C. (1986) DNA 5, 271.
- Houts, G.E., Miyagi, M., Ellis, C., Beard, A., and Beard, J.W. (1979) J. Virol. 29, 517.
- Kotewicz, M.L., Sampson, C.M., D'Alessio, J.M., and Gerard, G.F. (1988) Nuc. Acids Res. 16, 265.

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